

Determination of Phthalate Esters in Soft Drinks By GC-MS

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Key Words

TraceGOLD TG-5MS, phthalate esters, soft drinks, liquid-liquid extraction

Abstract

This application note demonstrates the quantitative analysis of phthalate esters in soft drinks. The combination of Thermo Scientific™ ultra low bleed TraceGOLD™ TG-5MS columns with a Thermo Scientific™ TRACE™ GC coupled with a Thermo Scientific™ ISQ™ mass spectrometer provides high sensitivity for the detection of phthalate esters in selected ion monitoring (SIM) acquisition mode.

Introduction

Phthalate esters are the main plasticizers used as softening agents in the production of PVC. These compounds are reported to act as endocrine disruptors, and exposure to high levels can cause harmful effects in the human reproductive system. There have been reports from the U. S. Food and Drug Administration that certain foods and beverages, particularly fruit juices, contain high levels of phthalates. In some cases, deliberate adulteration of soft drinks with phthalate esters has been reported.

The EU recently published a methodology without an extraction method for bis-(2-ethylhexyl)phthalate (DEHP) in sports drink at concentrations between 3000 and 100000 ng/mL [1]. In this application note, an analytical procedure for the quantitative analysis of 15 phthalate esters between the concentrations of 100 and 5000 ng/mL is reported. The extraction of the phthalate esters is based on liquid-liquid extraction. Extraction efficiencies are reported at low (300 ng/mL) and high (1000 ng/mL) concentrations in spiked drink sample.



Experimental Details

| Consumables | | Part Number |
|--------------------|---|-------------|
| Column: | TraceGOLD TG-5MS, 30 m × 0.25 mm × 0.25 μm | 26098-1420 |
| Septum: | Thermo Scientific BTO, 17 mm | 31303211 |
| Liner: | Thermo Scientific™ Splitless Straight Liner, 5 × 8 × 105 mm | 45350033 |
| Column ferrules: | 100% graphite ferrules for Thermo Scientific™ TRACE™ injector, 0.1–0.25 mm i.d. | 29053488 |
| Column ferrules: | Graphite/Vespel® for transfer line 0.1–0.25 mm i.d. | 29033496 |
| Injection syringe: | 10 μL fixed needle syringe for a Thermo Scientific™ TriPlus™ Autosampler | 36500525 |

| Sample Handling Equipment | | Part Number |
|---------------------------|---|-------------|
| System: | Thermo Scientific™ eVol™ Sample Dispensing System (containing Sample Dispensing System, eVol XCHANGE™ Syringes in 5, 50, and 500 μL volumes and eVol stand) | 66002-024 |
| Vials and closures: | Thermo Scientific 9 mm Wide Opening Screw Thread Vial Convenience Kit, 2 mL Clear Vial with Patch, Blue Polypropylene Closure with Clear PTFE/Blue Silicone Septa | 60180-599 |

| Chemicals and Reagents | | Part Number |
|--|--|-------------|
| Fisher Scientific™ HPLC grade acetone | | A/0600/15 |
| Fisher Scientific HPLC grade dichloromethane | | D/1856/17 |
| Fisher Scientific HPLC grade hexane | | H/0406/15 |
| Fisher Scientific HPLC grade water | | W/0106/17 |

Preparation of Calibration Standard

A stock standard solution of 1000 μg/mL of phthalate esters listed in Table 1 was prepared in hexane / acetone (8:2 v/v) (Standard A). This was then used to prepare standard solutions in hexane of 5000, 3000, 1000, 500, 300, and 100 ng/mL. These were Standards B–G. For construction of the calibration curve, the standards B–G were fortified with benzyl benzoate as an internal standard to a level of 1000 ng/mL.

Sample Preparation

The standards and extracts were prepared in scrupulously clean glassware. Avoiding any contact with plastic is vital as phthalates can contaminate glassware and blank samples very easily. Glass analytical syringes, glass pipettes, and pesticide grade solvents were used for preparing samples.

Glassware was scrupulously cleaned by rinsing first with water and then with acetone and hexane. For accuracy, the calibration samples were prepared using an eVol dispensing system. This avoided the use of plastic pipettes as the eVol system has a glass syringe barrel and stainless steel dispensing needle. Any plastic containers were avoided to reduce potential phthalate contamination in the sample preparation.

To prepare the spiked soft drink samples, 300 ng/mL and 1000 ng/mL of phthalate esters (listed in Table 2) prepared in acetone were spiked into 5 mL of soft drink followed by the addition of 5 mL of dichloromethane containing 1000 ng/mL internal standard. The solution mixture was shaken vigorously. An aliquot of organic layer was transferred to the GC vial.

A high injector temperature (320 °C) was used to release the high MW phthalates or any adsorbing phthalates in the injector head. The high temperature resistant BTO septa were used.

Separation Conditions

| | |
|-----------------------|---|
| Instrumentation: | Thermo Scientific™ TRACE™ GC Ultra gas chromatograph |
| Carrier gas: | Helium |
| Column flow: | 1.0 mL/min, constant flow |
| Oven temperature: | 80 °C (1 min), 10 °C/min, 320 °C (8 min) |
| Injector type: | Split/Splitless |
| Injector mode: | Splitless (1 min), 20 mL/min flow rate, constant septum purge |
| Injector temperature: | 320 °C |

MS Conditions

| | |
|----------------------------|--|
| Instrumentation: | Thermo Scientific™ ISQ™ GC single quadrupole mass spectrometer |
| Transfer line temperature: | 300 °C |
| Source temperature: | 260 °C |
| Ionization conditions: | EI |
| Electron energy: | 70 eV |
| SIM scan parameters: | Table 1 |

| Compound | Abbrev | Scan start time (min) | Quan ion (Qual Ion) | Dwell time (s) |
|--|--------|-----------------------|---------------------|----------------|
| Dimethylphthalate | DMP | 7.0 | 163 (194, 77) | 0.08 |
| Diethylphthalate | DEP | 10.50 | 149 (177, 121) | 0.08 |
| Benzyl benzoate (Internal standard) | ISTD | 12.50 | 105 (212, 91) | 0.08 |
| Diisobutyl phthalate | DIBP | 14.00 | 149 (223, 205) | 0.08 |
| Di-n-butyl phthalate | DBP | 15.00 | 149 (223, 205) | 0.08 |
| Bis(2-methoxyethyl) phthalate | DMEP | 15.75 | 59 (149,193) | 0.08 |
| Bis(4-methyl-2-pentyl) phthalate | BMPP | 16.30 | 149 (251,167) | 0.08 |
| Bis(2-ethoxyethyl) phthalate | DEEP | 16.80 | 45 72,149 | 0.08 |
| Dipentylphthalate | DPP | 17.20 | 149 (237, 219) | 0.08 |
| Di-n-hexyl phthalate | DHXP | 18.20 | 104 (149, 76) | 0.04 |
| Benzyl butyl phthalate | BBP | 18.20 | 149 (91, 206) | 0.04 |
| Bis(2-n-butoxyethyl) phthalate | DBEP | 19.50 | 149 (223, 205) | 0.08 |
| Dicyclohexyl phthalate | DCHP | 20.30 | 149 (167, 83) | 0.08 |
| Bis(2-ethylhexyl) phthalate | DEHP | 20.30 | 149 (167, 279) | 0.08 |
| Di-n-octyl phthalate | DNOP | 21.40 | 149 (279, 167) | 0.08 |
| Di-nonyl Phthalate | DNP | 22.80 | 149 (293, 71) | 0.08 |

Table 1: SIM Scan parameters

Injection Conditions

| | |
|-------------------------------------|---------------------|
| Instrumentation: | TriPlus Autosampler |
| Injection volume: | 1 µL |
| Pre- and post-injection dwell time: | 3 s |

Data Processing

| | |
|-----------|---------------------------------------|
| Software: | Thermo Scientific™ Xcalibur™ software |
|-----------|---------------------------------------|

Results

The analysis was carried out using a TraceGOLD TG-5MS column. The chromatogram of 1000 ng/mL of spiked phthalate ester in soft drink, in a full scan mode, is shown in Figure 1.

Phthalate esters are environmentally ubiquitous and this may therefore affect measured recoveries. To minimize this problem, care was taken when preparing solutions for calibration and extractions. Longer chain phthalate esters such as DNP, DNOP, and DHXP can adsorb to the glassware and the lower extraction recoveries were expected.

The results for the calibration data and for the phthalate esters extracted from a soft drink are displayed in Table 2. The coefficients of determination (R^2) between the area ratio of sample and internal standard for all phthalate esters were > 0.99 (Table 2), demonstrating excellent method linearity using the TraceGOLD TG-5MS GC column.

The extraction of phthalate esters at 300 ng/mL and 1000 ng/mL gave recoveries between 66–111% and 71–118%, respectively.

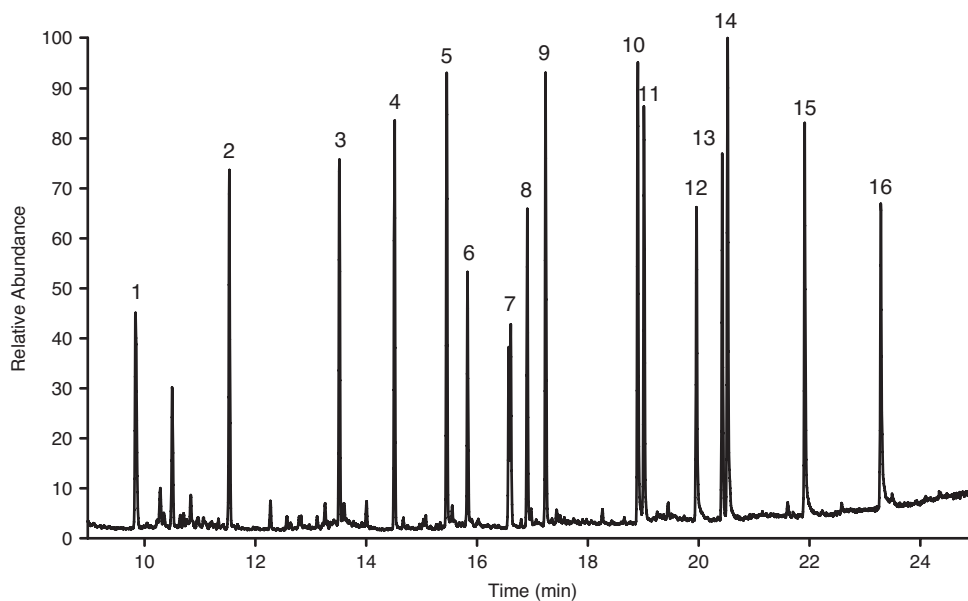


Figure 1: TIC of 1000 ng/mL phthalate esters standard in full scan 40–450 amu

| Peak | Compound | Abbrev | t_r (min) | R^2 | % Recovery at 300 ng/mL (n=3) | % Recovery at 1000 ng/mL (n=2) |
|------|-------------------------------------|--------|-------------|--------|-------------------------------|--------------------------------|
| 1 | Dimethylphthalate | DMP | 9.91 | 0.9944 | 103 | 85 |
| 2 | Diethylphthalate | DEP | 11.60 | 0.9953 | 106 | 108 |
| 3 | Benzyl benzoate (Internal standard) | ISTD | 13.60 | | | |
| 4 | Diisobutyl phthalate | DIBP | 14.58 | 0.9970 | 92 | 118 |
| 5 | Di-n-butyl phthalate | DBP | 15.52 | 0.9964 | 91 | 92 |
| 6 | Bis(2-methoxyethyl) phthalate | DMEP | 15.90 | 0.9918 | 93 | 96 |
| 7 | Bis(4-methyl-2-pentyl) phthalate | BMPP | 16.6, 16.68 | 0.9975 | 95 | 82 |
| 8 | Bis(2-ethoxyethyl) phthalate | DEEP | 16.98 | 0.9913 | 100 | 93 |
| 9 | Dipentylphthalate | DPP | 17.31 | 0.9940 | 107 | 87 |
| 10 | Di-n-hexyl phthalate | DHXP | 18.98 | 0.9945 | 84 | 71 |
| 11 | Benzyl butyl phthalate | BBP | 19.09 | 0.9937 | 111 | 86 |
| 12 | Bis(2-n-butoxyethyl) phthalate | DBEP | 20.03 | 0.9930 | 77 | 77 |
| 13 | Dicyclohexyl phthalate | DCHP | 20.50 | 0.9925 | 107 | 84 |
| 14 | Bis(2-ethylhexyl) phthalate | DEHP | 20.60 | 0.9941 | 100 | 78 |
| 15 | Di-n-octyl phthalate | DNOP | 21.99 | 0.9962 | 71 | 80 |
| 16 | Di-nonyl Phthalate | DNP | 23.37 | 0.9947 | 66 | 79 |

Table 2: Recoveries and linearity of phthalate esters according to their retention times

Conclusion

A method for the quantification of phthalate esters in soft drinks has been developed on a TraceGOLD TG-5MS column. The ultra low bleed characteristics of the column enable the detection of low levels of phthalate esters using the ISQ mass spectrometer in SIM mode.

References

- [1] EU commission JRC technical notes: Determination of bis-(2-ethylhexyl)phthalate (DEHP) in sports drinks by isotopic dilution headspace solid-phase micro extraction gas chromatography mass spectrometry (HS-SPME-GC-MS).

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